

Comparative study of protection in pigs immunised by different routes and doses with the naturally attenuated African swine fever virus isolate OUR T88/3 and role of immunomodulatory cytokines.

> Tamara Jabbar, David Chapman Pedro Sanchez-Cordon, Ana L. Reis, Lynnette Goatley, Christopher L. Netherton and Linda Dixon The Pirbright Institute, Ash road, Pirbright, Woking GU24 ONF, UK. Tel. +44 (0)1483 232441. Email: enquiries@pirbright.ac.uk www.pirbright.ac.uk

Introduction

African swine fever (ASF) is a devastating disease of domestic swine, with a mortality rate reaching 100% for highly pathogenic isolates.

The apparently unstoppable spread of AST throughout the trans-Caucasus region and Russia since its introduction in 2007, as well as the



recent incursions into Eastern Europe has highlighted the need for an effective vaccine to help control the disease.

- Control relies on rapid diagnosis, implementation of quarantine and slaughter of pigs.
- Despite the safety concerns raised by the use of a live attenuated vaccine (reversion to a virulent form or capability of causing subclinical or chronic ASF forms), to date, these vaccines have been the only form capable of conferring a reliable and effective protection against infections with homologous and occasionally heterologous virulent isolates of ASFV.

ABSTRACT





Comparative assessments were utilised to demonstrate differences between various combinations of doses (10³, 10⁴ and 10⁵ TCID50/ml), and routes of immunisation (intramuscular and intranasal) of low virulent African swine fever virus (ASFV) OURT88/3 in pigs.

Intranasal immunisations with low and moderate doses (10³ and 10⁴ TCID50/ml), provided complete protection (100%), mild and transient clinical signs and transient, moderate virus genome levels in blood against challenge with virulent genotype I OURT88/1 isolate.

In contrast, pigs immunised intramuscularly with low and moderate doses (10³ and 10⁴ TCID50/ml) displayed lower protection rates (50 to 66%). Nevertheless, protected animals showed no clinical signs, and only low or undetectable levels of virus genome in blood throughout the study.

Materials and Methods

Cells and viruses:

Results

Intranasal immunisations provided 100% protection in pigs vaccinated with low and moderate doses (10^3 and 10^4 TCID₅₀), and 66% protection in pigs

Conclusions

Intranasal immunisations with 10^3 and 10⁴ OURT88/3 provided 100% survival rate in vaccinated animals.

Viruses were grown and titrated in primary macrophage cultures. Titres are presented as (TCID50/ml).

Experimental design:

Animal experiments were conducted in BSL-3 facilities at CReSA (Barcelona, Spain), according to regulated procedures from the Animals (Scientific Procedures) Act 1986

| | | | Immunisation | Challenge by IM |
|------------|--------|-----------|--------------------------|--------------------------|
| Experime | No. of | Immunisat | dose of OUR | inoculation of OUR |
| ntal group | pigs | ion route | T88/3 | T88 /1 |
| | | | (TCID ₅₀ /ml) | (TCID ₅₀ /ml) |
| А | n=6 | IM | 10^{3} | 104 |
| В | n=6 | IN | 10^{3} | 10^{4} |
| С | n=6 | IM | 10^{4} | 10^{4} |
| D | n=6 | IN | 10^{4} | 10^{4} |
| E | n=6 | IM | 10^{5} | 10^{4} |
| F | n=6 | IN | 10^{5} | 10^{4} |
| G | - 2 | | | 1.04 |
| (Control) | n=3 | - | - | 10. |

IN: Intranasal immunisation; IM: intramuscular immunisation

• Sampling, clinical and post-mortem examination:

Rectal temperatures and clinical signs were monitored daily. EDTA blood and serum samples were collected on day (0, 3, 5, 7, 14 and 21) postimmunisation and day (3, 5, 7, 14 and 19) post-challenge.

vaccinated with high dose (10^5) of OURT88/3.

Intramuscular immunisations induced lower protection with a survival rate of **50%** and 66 % in pigs vaccinated with OURT88/3 (10^3 TCID_{50}) and (10^4 TCID₅₀) respectively



- Surviving animals (viraemic), had low virus titre on termination and no virus was detected in the tissues.
- Immunised animals that died, had high virus titres $(10^7 \text{ or } 10^8)$
- Protected pigs did not display changes in serum cytokine levels, whereas an increase of IL-10 and IFNy appeared post challenge in non-protected pigs with acute ASF.

(A)

g/m

(B)

1 ²⁰⁰ 1 m/b

- Survival of pigs after challenge was associated with a balance between pro- (TNF α and IL-1 β) and antiinflammatory cytokines (IL-10), without participation of IFNy.
- In contrast, animals that died showing an acute form of ASF displayed an imbalance linked to an exacerbated increase of IL-10 along antibody anomalous with an response.

Future work

- Further testing of vaccine candidates.
- Testing duration of immunity.
- **Testing onset of immunity.**

Non-protected pigs were euthanized at a specified endpoint, while protected pigs were euthanized at 19 dpc.

• ASFV detection and immune response evaluation:

Blood and tissues samples were analysed for ASFV genome detection by quantitative PCR (qPCR) (King et al., 2003).

Serum samples were assayed by commercial ELISA kits (Blocking ELISA), for detection of ASFV-specific antibodies against VP72 (INGEZIM PPA Compac, Ingenasa), or porcine immunoregulatory cytokines (TNFα, IFNγ, IL-4 and IL-10) (R&D Systems).



(A) IL-10









